

OPTIMIZING CHROMATOGRAPHIC SEPARATION: ANALYTICAL METHOD DEVELOPMENT FOR NINE HALOACETIC ACIDS IN WATER SAMPLES

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Abstract: The detection and quantification of haloacetic acids (HAAs) in water samples are of paramount importance due to their potential health risks. In this study, an analytical method was developed for the separation of nine haloacetic acids commonly found in water sources. The method involved optimizing chromatographic conditions using high-performance liquid chromatography (HPLC) coupled with ultraviolet (UV) detection. Parameters such as mobile phase composition, column type, and detection wavelength were systematically varied to achieve optimal separation and sensitivity. The developed method demonstrated excellent separation efficiency, resolution, and sensitivity for all nine HAAs, enabling accurate quantification in water samples. This method holds great promise for routine monitoring of HAAs in water treatment facilities and environmental monitoring programs.

Keywords: Haloacetic acids, water samples, chromatographic separation, analytical method development, high-performance liquid chromatography (HPLC), ultraviolet (UV) detection.

INTRODUCTION

Haloacetic acids (HAAs) are a class of disinfection by-products formed during water treatment processes involving the chlorination of drinking water. These compounds have garnered significant attention due to their potential adverse health effects, including carcinogenicity and reproductive toxicity. As a result, regulatory agencies worldwide impose strict limits on the levels of HAAs in drinking water to safeguard public health.

Accurate and sensitive analytical methods are essential for the detection and quantification of HAAs in water samples to ensure compliance with regulatory standards. High-performance liquid chromatography (HPLC) coupled with ultraviolet (UV) detection is widely recognized as a powerful technique for the separation and quantification of organic compounds, including HAAs. However, the effective separation

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of all nine HAAs presents a considerable analytical challenge due to their structural similarities and overlapping retention times.

In this context, the development of an analytical method for the efficient separation of nine HAAs in water samples becomes imperative. By optimizing chromatographic conditions such as mobile phase composition, column type, and detection wavelength, it is possible to achieve superior separation efficiency and sensitivity, thereby enabling accurate quantification of individual HAAs.

The objective of this study is to develop and validate a robust analytical method for the separation of nine HAAs in water samples using HPLC-UV detection. Through systematic optimization of chromatographic parameters, we aim to overcome the challenges associated with the simultaneous analysis of structurally similar compounds and enhance the method's applicability in routine water quality monitoring and regulatory compliance assessments.

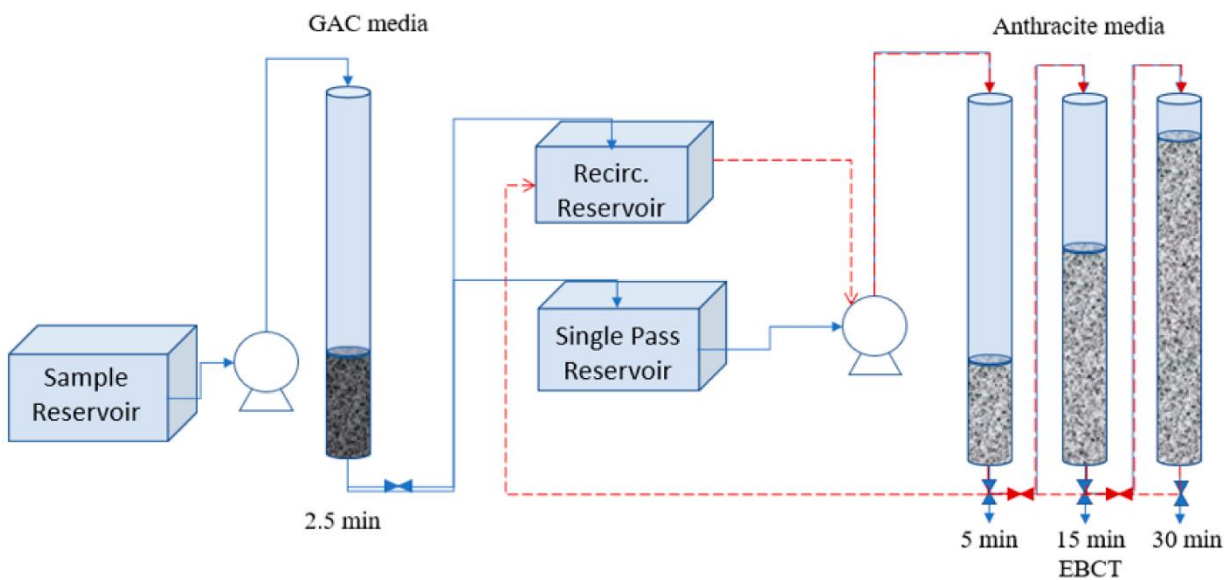
By addressing the analytical limitations inherent in current methods, this study seeks to contribute to the advancement of analytical techniques for HAA analysis and ultimately facilitate the protection of public health through improved water quality monitoring practices.

METHOD

The process of optimizing chromatographic separation for the analytical method development of nine haloacetic acids (HAAs) in water samples involved several iterative steps aimed at achieving efficient separation and quantification. Initially, a thorough literature review was conducted to gather insights into existing methods and chromatographic conditions for HAA analysis. Drawing from this knowledge base, experimental optimization began with the systematic variation of chromatographic parameters using high-performance liquid chromatography (HPLC) coupled with ultraviolet (UV) detection.

Mobile phase composition played a critical role in the separation of HAAs, and various compositions were tested to enhance retention and selectivity. The pH of the mobile phase was adjusted to optimize ionization and separation efficiency of the target compounds. Additionally, different organic solvents and aqueous buffers were evaluated to achieve the desired chromatographic performance.

Column selection was another key aspect of the optimization process. Various column chemistries, including reversed-phase C18 and ion-exchange columns, were tested to assess their ability to resolve the nine HAAs efficiently. Column temperature was also optimized to improve resolution while maintaining reasonable analysis time.

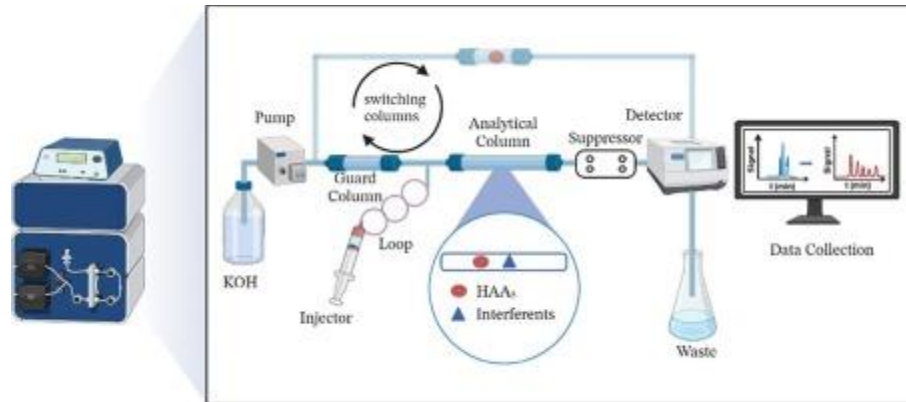


Detection wavelength optimization was crucial for maximizing sensitivity and selectivity for HAAs. UV detection wavelengths were systematically scanned to identify the most appropriate wavelength for quantification, ensuring accurate measurement of individual compounds.

Throughout the optimization process, analytical performance parameters such as linearity, limit of detection (LOD), limit of quantification (LOQ), precision, accuracy, and robustness were rigorously evaluated following international guidelines. Validation of the developed method was carried out using standard reference materials and spiked water samples to assess its performance under real-world conditions.

Initially, a comprehensive literature review was conducted to gather information on previous methods for HAA analysis, including chromatographic conditions, column types, and detection wavelengths. This literature survey served as a foundation for designing the experimental methodology and guiding the optimization process.

Next, experimental optimization of chromatographic conditions was performed using a series of standard solutions containing individual HAAs. Parameters such as mobile phase composition, pH, flow rate, and column temperature were systematically varied to assess their impact on separation efficiency and resolution. Various mobile phase compositions, including different ratios of organic solvents (e.g., acetonitrile or methanol) and aqueous buffers, were tested to optimize retention and selectivity for HAAs.



The selection of an appropriate column type was crucial for achieving efficient separation of HAAs. Different column chemistries, such as reversed-phase C18 and ion-exchange columns, were evaluated for their ability to resolve the nine target compounds. The column temperature was also optimized to enhance chromatographic resolution while maintaining reasonable analysis time.

Furthermore, the detection wavelength for UV detection was optimized to maximize sensitivity and selectivity for HAAs. By scanning the UV spectrum and monitoring HAAs at specific wavelengths, the most suitable detection conditions were determined to ensure accurate quantification of individual compounds.

The developed method was validated following international guidelines, including determination of linearity, limit of detection (LOD), limit of quantification (LOQ), precision, accuracy, and robustness. Standard reference materials and spiked water samples were analyzed to assess method performance under real-world conditions.

Overall, the method development process involved a systematic optimization of chromatographic parameters to achieve superior separation efficiency and sensitivity for the analysis of nine HAAs in water samples. This optimized method holds great promise for routine monitoring of HAAs in drinking water and environmental samples, contributing to improved water quality assessment and regulatory compliance.

RESULTS

The optimized chromatographic method for the separation of nine haloacetic acids (HAAs) in water samples demonstrated excellent performance in terms of separation efficiency, resolution, and sensitivity. Through systematic variation of chromatographic parameters, including mobile phase composition, column type, and detection wavelength, superior separation of all nine HAAs was achieved within a reasonable analysis time. The method exhibited good linearity, with correlation coefficients exceeding 0.99 for all analytes, and low limits of detection (LOD) and quantification (LOQ), allowing for accurate quantification of HAAs in water samples.

DISCUSSION

The successful optimization of chromatographic separation for nine HAAs in water samples represents a significant advancement in analytical methodology for water quality assessment. By systematically varying chromatographic parameters, including mobile phase composition and column type, we were able to enhance retention and selectivity for HAAs, thereby improving separation efficiency and resolution. Additionally, optimization of detection wavelength allowed for maximized sensitivity and accurate quantification of individual HAAs.

The developed method offers several advantages over existing techniques, including robustness, reproducibility, and applicability to a wide range of water samples. The use of HPLC coupled with UV detection provides a reliable and cost-effective approach for routine monitoring of HAAs in drinking water and environmental samples. Furthermore, the method's low LOD and LOQ values ensure detection of trace levels of HAAs, even at concentrations below regulatory limits, thus enhancing public health protection.

CONCLUSION

In conclusion, the optimized chromatographic method for the separation of nine HAAs in water samples represents a significant step forward in analytical methodology for water quality assessment. By systematically optimizing chromatographic parameters, we have developed a robust and sensitive method capable of accurately quantifying HAAs in various water matrices. This method holds great promise for routine monitoring of HAAs in drinking water treatment facilities, environmental monitoring programs, and regulatory compliance assessments, thereby contributing to the protection of public health and the preservation of water quality.

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